Introduction

Streptococcus agalactiae or Group B Streptococcus (GBS) is gram-positive coccus which appears in chain or pairs. The organism is found as commensal in the gastrointestinal and the genitourinary tract of up to 30% of healthy adults, GBS is facultative anaerobic, catalase-negative gram-positive cocci, oxidase negative. Their metabolism is mainly fermentative and lactic acid is the predominant end product [1].

Streptococci may display beta-hemolytic, alpha-hemolytic and non-hemolytic reactions on blood agar. Those with beta-hemolytic ability, i.e. to lyse erythrocytes completely in blood agar, were subdivided by their reaction to specific antisera against their group-specific cell wall anchored carbohydrate [2]. GBS described as zoontic organisms, It is a member of the normal flora of the female genital tract, also could be isolated from bovine milk or from vaginal and rectal swab specimens from asymptomatic pregnant women[3].

Many of the prevalence studies looking at colonization have been carried out
on pregnant women and from these it is known that colonization rates are higher in sexually active individuals, especially those with multiple sexual partners, and increases with maternal age [4].

The ability of GBS to cause infections is multifactorial. The main virulence associated GBS proteins identified to date are secreted or surface components, including β-haemolysin|cytolysin operon (cyl-operon) which promote enhancing bacterial pathogenicity.

The GBS has β- Hemolysin\Cyto
tysin operon which considered as surface-associated toxins that play an important role in virulence of the organism, also this is responsible for the characteristic zone of clearing around GBS colonies grown on blood agar plates[5].

One of the most virulence factors for Streptococcus agalactiae that mediates in wall anchoring are sortase enzymes which have a major role in some diseases such as meningitis and neonatal sepsis.

Among these enzymes are sortase A which is considered as a housekeeping sortases and was shown experimentally to anchor a large number of surface proteins[6].

In addition to sortases A, there are two other specialized sortases, one associated with (fibronectin–binding , collagen binding-t-antigen) refers to sortase B . While the other recognizes an altered consensus sequence, which is called sortase C [7].

**Patients and Methods**

**Patients:**
This study included 84 women patients suffering from vaginitis who were submitted to Hilla Teaching Hospital during a period of three months from October to January 2013.

**Collection of specimens:**
Specimens were collected from patients with vaginitis. The swabs were inserted into the upper part of the vagina and rotated there before withdrawing it, so that exudates was collected from the upper as well as the lower vaginal wall. An endocervical swab must be collected. A vaginal speculum must be used to provide a clear sight of the cervix and the swab was rubbed in and around the introitus of the cervix and withdrawn without contamination from the vaginal wall.

Swabs for culture were placed in tubes containing normal saline to maintain the swab moist until taken to laboratory. The swab has been inoculated on Colombia agar media and incubated aerobically for 24hr. at 37°C.

**Detection of Sortase Enzymes (A&C1) & Cyl Operon (A& B ) by PCR.**
Bacterial DNA extract was used as a template in specific PCRs for the detection of virulence genes (Str A, Str C1 ,Cyl A & B). DNA was purified from bacterial cells by using the wizard minipreps DNA Kit(Geneaid-USA). A primer were used for the amplification of a fragment genes according to the references mentioned in tables (1) and (2).
### Table 1: Primers sequences and PCR conditions to detect Sortase genes.

<table>
<thead>
<tr>
<th>Genes</th>
<th>Primer sequence (5’-3’)</th>
<th>Size of product bp</th>
<th>PCR condition</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Srt A F</td>
<td>TACTTCCAATCCAATGCATCTGCTCAAAACGAAATCACAATAAATGCTTTTAATATCGACTCAT</td>
<td>500</td>
<td>94°C 3min 1x</td>
<td>[8]</td>
</tr>
<tr>
<td>Srt A R</td>
<td></td>
<td></td>
<td>94°C 2min</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>63°C 1min 28x</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>72°C 1min</td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>72°C 10min 1x</td>
<td></td>
</tr>
<tr>
<td>Srt C1 F</td>
<td>AAAGGATCCCTCTCACGCCAATATTAATGCTT</td>
<td>370</td>
<td>95°C 2.5min 1x</td>
<td>(8)</td>
</tr>
<tr>
<td>Srt C1 R</td>
<td>AAAGGATCCCTATTTGTGCTGCTGAAGGTCTT</td>
<td></td>
<td>94°C 30sec</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>53°C 1min 30x</td>
<td></td>
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<td></td>
<td>72°C 30sec</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>72°C 7min</td>
<td></td>
</tr>
</tbody>
</table>

### Table 2: Primers sequences and PCR condition to detect Cyl A & B by PCR.

<table>
<thead>
<tr>
<th>Genes</th>
<th>Primer sequence (5’-3’)</th>
<th>Size of product bp</th>
<th>PCR condition</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cyl A F</td>
<td>GACTCGGGGATTGATAGGC</td>
<td>688</td>
<td>95°C 5min 1x</td>
<td>[9]</td>
</tr>
<tr>
<td>Cyl A R</td>
<td>GCTGCTAAAGCTGCCTTAC</td>
<td></td>
<td>94°C 1min 1x</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>58°C 1min 28x</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>72°C 1min</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>72°C 10min 1x</td>
<td></td>
</tr>
<tr>
<td>Cyl B F</td>
<td>CGCCGCGGATCCATATTATTGGCTCATTTGAGG</td>
<td>428</td>
<td>94°C 5min 1x</td>
<td>[10]</td>
</tr>
<tr>
<td>Cyl B R</td>
<td>CGCCGCGGATCCATATTATTGGCTCATTTGAGG</td>
<td></td>
<td>94°C 1min 1x</td>
<td></td>
</tr>
<tr>
<td></td>
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<td></td>
<td>56°C 1min 30x</td>
<td></td>
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<td></td>
<td>72°C 1min</td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>72°C 10min 1x</td>
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</table>
Results and Discussion
Bacterial isolates:
Only 20 isolates of GBS were obtained in this study which have been isolated from pregnant and non-pregnant women. All isolates have been cultured in Colombia agar media and incubated aerobically for 24hr. at 37°C.

Detection of sortase A & C1 enzyme by PCR:
Results found that sortase A was observed in 7 isolates of GBS strains (Figure 1). Its well known that sortase A is housekeeping enzyme that may be present in all Streptococcus strains especially St. agalactiae and St. pyogens in local study, so this results confirms that sortase A is normally present in this bacteria as a result of its being housekeeping [11].

The Gram positive bacteria including Streptococcus agalactiae assemble pili by distinct mechanism involving a trans peptidase called sortase, regarding to the others , it was found that sortase A was exist in all GBS isolates, this result could prove that sortase A is housekeeping enzyme which involved not only information of pili type sortase, but also anchors pili to the cell wall [12].

![Ladder 1 2 3 4 5 6 7](image)

**Figure 1** Gel electrophoresis of PCR product of sortase (A) *(1, 2, 3, 4, 5, 6, 7) GBS isolates with positive result for sortase A

This result is in agreement with the results obtained by other studies [13]. Moreover, Srt C gene which encodes sortase C enzyme, class C sortases are integral membrane cysteine transpeptidases of gram-positive bacteria. Sortase C1 was identified in this study (SrtC1) in three isolates of GBS as shown in figure (2).

![500 bp Ladder](image)

This result is in agreement with results are obtained by other studies where it was reported that this enzyme are present in GBS and responsible for invasive infection in non pregnant adults [14]. Sortase C1, not as in sortase A, is not classified as housekeeping enzyme, so it's not ubiquitous as sortase A.
According to data obtained in this study, it was observed that only three isolates have been found to contain all Srt genes and this will increase the anchored product.

**Detection of Cyl A & B by PCR:**
Molecular detection of cytolycin operon was done using haemolycin primer of *Streptococcus agalactiae*. It was found that cyl A was observed in all GBS strains as shown in figure (3), while cyl B was found only in three isolates as shown in figure (4).

**Figure 2** Gel electrophoresis of PCR product of sortase (C1) produced by *Streptococcus agalactiae* *(2, 3, 4)* positive for sortase C1, *(1, 5, 6, 7)* negative for sortase C1

Specifically, SrtA cleaves the target protein between the threonyl and glycyl residues of the LPXTG motif to form an acyl enzyme intermediate. This is then resolved by the nucleophilic attack of amino groups, usually provided by the lipid II precursor of peptidoglycan, which is subsequently incorporated into the cell-envelope. By contrast, the SrtC enzymes of GBS are located within the PIs, and function to polymerize only those LPXTG proteins located within the same PI *(15)*.

**Figure 3** Gel electrophoresis of PCR product of *S. agalactiae cyl A* operon *(1, 2, 3, 4, 5, 6, 7)* GBS isolates with positive result
Figure 4 Gel electrophoresis of PCR product of S.agalactiae cyl B *(4, 5, 6,7) GBS isolates with negative result

These result are correlated with results obtained by other studies where it was found that the ABC transporter encoded by this operon (cylA and cylB) was proposed to be required for hemolysin export at the bacterial surface [16]. The β-hemolysin represents an important virulence factor of St. agalactiae, its ability to damage erythrocytes, lung epithelial cells, and brain microvascular endothelial cells is regarded as an initial step in invasive disease. It was found that the genes cylA and cylB display homology to typical ABC transporters, with Cyl A as the ATP-binding domain and Cyl B as the transmembrane protein [17].

Also, other results were obtained by others, who observed that the β-hemolysin/cytolysin expressed by GBS is an important virulence factor encoded within a cluster of twelve genes forming the cyl operon, hemolytic activity of GBS is always associated with the synthesis of an orange pigment. [18]

The deduced amino acid sequences of cylA and cylB demonstrate significant homologies to prokaryotic and eukaryotic ABC transporters. The typical ABC transporter consists of two transmembrane domains and two highly conserved ATP-binding domains [19]. The transmembrane domains of ABC transporters are highly hydrophobic, and the majority of transporters are predicted to have six membrane-spanning segments per domain. Cyl B codes for a hydrophobic protein with six potential membrane-associated segments.

Both genes, cylA and cylB, appear to be required for the hemolytic activity of S. agalactiae wild-type strains, since several naturally occurring non-hemolytic S. agalactiae strains harbor an insertion sequence in the cylA gene and targeted mutants of cylB exhibit a non-hemolytic and non-pigmented phenotype that propose that CylA and CylB constitute a typical ABC transporter catalyzing an ATP-dependent export of hemolysin and pigment.

Both CylA and CylB are transporters, they are not responsible for synthesis of hemolysin because other Cyl genes may play a role in production of Hly. Such CylE is located downstream to CylB. However, the absence of CylB from our isolates does not have an effect on Hyl production because CylB is just...
transmembrane protein that Cyl A may give the same function.

**Conclusions**
The study has arrived at the following conclusions:

1. Only twenty isolates of GBS were obtained in spite of all age groups of patients were included.
2. Sortase enzymes A, and C were detected in some GBS isolates which play a role in anchoring the surface proteins.
3. Cyl operon A and B also were detected in some GBS isolates which play a role in ATP-binding domain (ABC-Transporter cassette).

**Acknowledgements**
I would like to express my thanks to "Allah" and his prophet "Mohammad."

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**References:**